

# **The nuclear import receptor Kpn $\beta$ 1 and its potential as an anti-cancer therapeutic target**

**Pauline J. van der Watt, Catherine L. Stowell and Virna D. Leaner\***

Division of Medical Biochemistry, Institute of Infectious Disease and Molecular Medicine,  
Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa

Keywords: Kpn $\beta$ 1, nuclear transport, cancer, therapeutic target

\*Correspondence to:

Dr Virna Leaner

Division of Medical Biochemistry

Faculty of Health Sciences

University of Cape Town

Observatory, Cape Town, 7925

South Africa

Ph: 27 21 406 6250

Fax: 27 21 406 6061

Virna.Leaner@uct.ac.za

## **Abstract**

Many proteins require transport across the nuclear envelope, the physical barrier separating the nucleus from the cytoplasm. Karyopherin  $\beta$  proteins are the major nuclear receptor proteins in the cell that cargo proteins across the nuclear envelope, allowing them to enter and exit the cell nucleus. Karyopherin  $\beta$ 1 (Kpn $\beta$ 1), a major nuclear import receptor, plays an integral role in importing transcription factors, cell signaling proteins, cell cycle proteins, etc. into the nucleus, thus playing a crucial role in maintaining normal cell homeostasis. However, cancer cells appear to differentially regulate the expression of the Karyopherin  $\beta$  proteins, presumably in order to maintain increased nuclear transport rates, thus implicating this protein family as a target for cancer therapy. The role of Kpn $\beta$ 1 in cancer is only now being elucidated, and recent work points to its potential usefulness as an anti-cancer target.

## **Introduction**

Access to the nucleus is limited by the nuclear envelope, a double-membrane structure in which large pore structures named nuclear pore complexes (NPC) are embedded. Molecules smaller than approximately 20-30 kDa can passively diffuse through the NPC, however, larger macromolecules generally require active receptor-mediated transport for entry/exit into the nucleus. The Karyopherin superfamily of proteins are soluble nuclear transport receptor proteins that are involved in the shuttling of cargo proteins, and certain RNAs, across the nuclear pore complex (NPC) into and out of the cell nucleus. The channel of the NPC through which Karyopherin-mediated transport occurs appears to be flexible and can

expand to allow for the transport of large cargoes. Each NPC is composed of up to around 100 different proteins, called nucleoporins (Nups) (1). The majority of Nups contain tandemly repeated phenylalanine-glycine (FG) motifs that line the central channel of the NPC. These are the motifs that Karyopherin proteins bind as they transport their cargoes through the pore.

The Karyopherin protein family is divided into alpha and beta subfamilies, with almost all known nuclear import and export pathways being mediated by Karyopherin  $\beta$  family members, while the Karyopherin  $\alpha$  family members are described as adaptor proteins (2). Although not all proteins needing to enter or exit the nucleus will utilise the Karyopherin proteins for transport, for example  $\beta$ -catenin translocates across the NPC on its own (3), a vast proportion of proteins within the cell depend on Karyopherin protein function, highlighting the importance of this protein family in maintaining correct cell functioning.

This review focuses on recent work that highlights the role of Karyopherin  $\beta$  proteins in cancer. We describe Karyopherin  $\beta$ -mediated transport, with specific focus on the association between Karyopherin  $\beta$  protein expression and cancer, and new inhibitors of the Karyopherin  $\beta$  proteins, specifically Kpn $\beta$ 1.

### **Karyopherin $\beta$ proteins as mediators of nuclear transport**

In mammalian cells there are more than twenty Karyopherin  $\beta$  family members. While some have been well characterised, the function of others is still unknown. Nine have been shown to act as export receptors (exportins), binding their cargo in the nucleus and transporting it through the NPC into the cytoplasm. Crm1 (the chromosome region maintenance 1 protein) is the major export receptor, also known as Exportin 1 (Xpo1). Crm1 recognises proteins containing a nuclear export signal (NES). Other Karyopherin  $\beta$  members are known to function as import receptors (importins); these proteins bind their cargo in the cytoplasm and transport it through the NPC into the nucleus. An import function has been demonstrated for eleven Karyopherin  $\beta$  family members. Although nuclear transport mediated by members of the Karyopherin  $\beta$  protein family is thought to occur by essentially similar mechanisms, the different receptors do appear to have distinct functions. Transportin 1, as an example, mediates the import of mRNA-binding proteins, while Importin 5 and Importin 7 mediate the import of ribosomal proteins (4). Moreover, some of the Karyopherin  $\beta$  members, such as Importin 13 and Exportin 4, are able to function as both import and export receptors. While diverse, there are certain features that characterise the Karyopherin  $\beta$  proteins; these include an N-terminal RanGTP-binding motif (RanGTP binding provides the energy required for Karyopherin  $\beta$ -mediated transport), a large size of between 90 and 130 kDa, an acidic isoelectric point of between 4.6 and 5.9, and an ability to interact directly with the nuclear pore complex (1;4).

### **Karyopherin $\beta$ 1 (Kpn $\beta$ 1), a major nuclear import receptor**

Kpn $\beta$ 1 (also known as importin  $\beta$ , p97, or yeast Kap95) was the first identified nuclear transport factor and is found at a concentration of approximately 3  $\mu$ M in the cell (5). It is

the major import receptor (importin protein) in the cell and can function in combination with up to 11 different partners to ferry distinct cargoes (2). Karyopherin  $\alpha$  family members (Kpn $\alpha$ 1-6) commonly act as adaptor proteins and work in concert with Kpn $\beta$ 1 to transport cargo proteins into the nucleus, although Kpn $\beta$ 1 has also been shown to partner with other Karyopherin  $\beta$ -like receptors. Each Kpn $\beta$ 1 heterodimer has distinct cargo specificities and the particular recognition signal on the cargo protein, as well as the general protein context in the cell, specify which adaptor protein should bind (6;7). While Kpn $\beta$ 1 commonly transports cargoes in concert with an adaptor, it can transport certain cargoes on its own; examples of this include transport of the parathyroid hormone-related protein (PTHrP) (8) and the sterol regulatory element binding protein 2 (SREBP-2) (9). The structure of Kpn $\beta$ 1 allows for its interaction with its many binding partners and cargoes, whereby the protein is coiled into a short superhelix, with extensive interaction surfaces both on the outside and the inside of the short superhelix (2). The Kpn $\beta$ 1 superhelix is comprised of 19 helical repeats, termed HEAT domains, each of which consists of two alpha helices joined by a loop.

Generally, Kpn $\beta$ 1 cargo proteins contain a specific recognition signal sequence or nuclear localisation signal (NLS) in their amino acid sequence. The first identified NLS was that of the SV40 large T-antigen (10), which consists of a short stretch of basic amino acids, designated as the basic type or “classical” NLS (cNLS). This type of NLS is divided into two groups, monopartite and bipartite (11). Monopartite NLSs contain a single cluster of basic lysine and arginine amino acids (PKKKRKV) (P: proline; K: lysine; R: arginine; V: valine), while bipartite sequences contain two clusters of basic residues separated by an unconserved linker region (KRX<sub>(10-12)</sub>KRRK) (11). The classical NLS-containing proteins are recognised by Kpn $\alpha$ , which

subsequently binds Kpn $\beta$ 1, through its importin  $\beta$ -binding domain (IBB), and classical nuclear import occurs (12). Not all import cargoes, however, contain a cNLS and many non-classical NLS motifs exist that do not require an adaptor protein like Kpn $\alpha$  for nuclear import, but rely on Kpn $\beta$ 1 directly, or alternatively one of the other Karyopherin  $\beta$  members.

### **Nuclear transport via Karyopherin $\beta$ 1 requires the Ran-GTP nuclear transport cycle**

Kpn $\beta$ 1-mediated nuclear transport requires energy provided by Ran, a small Ras-like guanosine triphosphatase (GTPase). In the classical nuclear import pathway, the cargo protein is recognised by Kpn $\alpha$  in the cytoplasm, becomes bound by Kpn $\beta$ 1, and localises to the nuclear envelope. After translocation through the nuclear pore complex, RanGTP binds, inducing the allosteric destabilisation and release of the import cargo and Kpn $\alpha$ . Kpn $\alpha$  is recycled back to the cytoplasm via its own nuclear exporter, the cellular apoptosis susceptibility protein, Cas (Cse1L). Kpn $\beta$ 1 is separately transported back to the cytoplasm, bound by RanGTP. Back in the cytoplasm, RanGTP is hydrolysed to RanGDP and Kpn $\beta$ 1 is released for its next cycle. RanGDP is transported back into the nucleus via its import factor, Ntf2 (13). Ran effectors are responsible for the GDP/GTP-state that Ran is in. Ran in the cytoplasm becomes bound by GDP due to the action of the Ran GTPase activating protein (RanGAP), as well as the Ran-Binding Protein 1 (RanBP1), localised in the cytoplasm. Ran in the nucleus, however, is in a GTP-bound state due to the RanGTP exchange factor (RanGEF or RCC1) that is bound to chromatin in the nucleus.

The structure of Kpn $\beta$ 1 supports its function, where HEAT repeats 1-8 (or residues 1-364) of the Kpn $\beta$ 1 protein are required for the binding of Kpn $\beta$ 1 to RanGTP, while the importin- $\beta$ -binding (IBB) domain of Kpn $\alpha$  interacts predominantly with residues 331-876 located within HEAT repeats 7-19 of Kpn $\beta$ 1 (5). Thus, RanGTP-mediated release of Kpn $\alpha$  from Kpn $\beta$ 1 is likely to be an active displacement rather than due to simple competition between Ran and Kpn $\alpha$  for a common binding site. In addition, Kpn $\beta$ 1 has at least two non-overlapping sites of interaction with the NPC (FG repeat binding sites), which could potentially be used sequentially during translocation (5). These are located between residues 152 and 352, corresponding to HEAT repeats 4-8 (14). It has been proposed that the two FG repeat binding sites could allow Kpn $\beta$ 1 to bind to two nuclear pore proteins at once, binding one and releasing the other as part of a multistep translocation through the nuclear pore (15).

### **Kpn $\beta$ 1 cargo molecules**

Kpn $\beta$ 1 imports numerous proteins that require entry into the nucleus. Multiple transcription factors are transported by Kpn $\beta$ 1, for example NF $\kappa$ B p65 (16) and Stat3 (17). These proteins are translated in the cytoplasm but are only active once in the nucleus and so depend on import into the nucleus following their translation for their function of binding and activation or repression of the promoters of target genes. Moreover, many cell signalling networks are highly dependent on the timely translocation of proteins from the cytoplasm into the nucleus. Extracellular signals bind to cell surface receptors, initiating cascades of protein phosphorylation and dephosphorylation events, leading to the nuclear translocation of various kinases, phosphatases, or transcription factors. Many of such signalling networks

depend on Kpn $\beta$ 1 protein function, for example the MAPK pathway, where phosphoERK enters the nucleus via Kpn $\beta$ 1 (18). In addition, cell cycle proteins are shuttled into or out of the nucleus when the appropriate signals are given (19). Kpn $\beta$ 1 thus plays a crucial role in the correct expression of genes, in cell signalling cascades, and in the control of the cell cycle, and in this way impacts many of the integral processes in the cell.

### **Additional Kpn $\beta$ 1 functions**

In addition to its nuclear transport functions, Kpn $\beta$ 1 is also an important regulator of mitosis, contributing to spindle assembly. After the breakdown of the nuclear envelope in mitosis, it has been shown that Kpn $\beta$ 1 regulates spindle assembly by binding to and sequestering key spindle assembly factors (SAFs), impairing their microtubule-stabilising or – organising activities (20). However, Kpn $\beta$ 1 is released from the SAFs in areas where RanGTP is present. RanGTP is produced only in the region of the mitotic chromosomes because the RanGEF RCC1 remains chromosome-bound at mitosis. Thus, near the mitotic chromosomes, RanGTP releases Kpn $\beta$ 1 from the SAF spindle assembly factors, allowing it to promote localised spindle formation.

Kpn $\beta$ 1 has also been implicated in a variety of other cellular processes including postmitotic nuclear envelope assembly and nuclear pore complex assembly (21). In addition, Kpn $\beta$ 1 has been implicated as a cytoplasmic chaperone, whereby it shields the basic domains of certain proteins (for example ribosomal proteins, histones and other basic nuclear proteins) preventing their aggregation with polyanions in the cytoplasm after translation (2). A

specific role has been shown for the Kpn $\beta$ 1/Imp7 heterodimer in suppressing the aggregation of histone H1 (22).

### **Regulation of Kpn $\beta$ 1 expression**

Since Kpn $\beta$ 1 is involved in multiple nuclear import pathways, as well as having several other non-import functions, of which new functions are likely to yet be elucidated, Kpn $\beta$ 1 literally impacts thousands of proteins and is crucial for the maintenance of correct cellular physiology (2). As normal cell homeostasis depends on the integrity of Kpn $\beta$ 1 function, its tight regulation and correct expression is very important. To date there are no known disease-causing mutations in Kpn $\beta$ 1, or any other components of the nuclear transport machinery (23). The absence of mutations and chromosomal translocations underscores the essential nature of Kpn $\beta$ 1 even at the single cell level.

Little is known regarding the regulation of Kpn $\beta$ 1 expression, although Kpn $\beta$ 1 expression has been shown to be responsive to stress, including heat shock, ethanol and oxidative stress (24). In addition, evidence has suggested that Kpn $\beta$ 1 expression is controlled in a cell cycle-dependent manner (in the yeast *Saccharomyces cerevisiae*) (25), likely impacting nuclear import rates during the cell cycle (26). A recent study by van der Watt et al. (2009) showed that the Kpn $\beta$ 1 promoter is regulated by the direct binding of the E2F transcription factor, a well known S-phase regulator, to three specific sites located in its promoter, verifying its cell cycle-dependent regulation (27). Moreover, in an *in silico* study performed by Quan et al. (2008), it was suggested that most Karyopherin  $\beta$  proteins, including Kpn $\beta$ 1, are regulated by Sp1, NFY, NRF-2 and RREB-1 transcription factors, which are all known to play important

roles in the cell cycle (26). Expression of Crm1, the major nuclear export receptor, has indeed been shown to be regulated by the binding of Sp1 and NFY transcription factors to its promoter (28).

The cell cycle regulation of Kpn $\beta$ 1 suggests that its expression may associate with proliferation. Indeed Kpn $\beta$ 1, along with twelve other Karyopherin  $\beta$  genes, shows comparatively high expression in tissues that proliferate actively, for example lymphocytes, tumours, testis and stem cells (23). Quan et al. (2008) state “the functional importance of  $\beta$ -karyopherins determines their expression” (26). Hence it is likely that Kpn $\beta$ 1 expression is increased where there is an increased requirement for nuclear import. In line with this, it has been shown that the rate of nuclear import can be modulated at least 10-fold by the concentration of Kpn $\beta$ 1 in the cell (29).

### **An association between nuclear transport proteins and cancer**

Cancer cells display altered nuclear transport. It is reported that as cells pass from quiescence to proliferating to transformed, the rates of nuclear transport increase (30;31). It is likely that nuclear transport rates are increased in transformed and cancer cells in order to sustain their increased metabolic and proliferative abilities. Kuusisto et al. (2011) recently reported that in order to increase nuclear transport rates, transformed cells display increased levels of certain Karyopherin  $\beta$  proteins (32). In line with this, over the last few years several lines of evidence have emerged supporting the idea that the increased nuclear transport rates in cancer are due, in part, to increased Karyopherin  $\beta$  protein expression. For

example, in several cancer cell lines and human tumours, the cellular apoptosis susceptibility protein (Cas or Cse1L), a Karyopherin  $\beta$  protein that mediates the nuclear to cytoplasmic translocation of Kpn $\alpha$ , is found overexpressed (33-38). This upregulation triggers the nuclear accumulation of Kpn $\alpha$ -dependent transcription factors, thereby mediating increased cell proliferation. In addition, high Crm1 expression has also recently been linked to cancer. Its overexpression has been reported in cervical cancer (39), ovarian cancer (40), osteosarcoma (41), glioma (42) and pancreatic cancer (43), with high levels of Crm1 being found to associate with poor patient survival. Similarly, overexpression of the Kpn $\beta$ 1 adaptor protein, Kpn $\alpha$ 2, has been reported in melanoma (44), breast cancer (45;46), cervical cancer (39), oesophageal cancer (47), ovarian cancer(48), non-small cell lung cancer (49), prostate cancer (50), bladder cancer (51) and liver cancer (52), and found to correlate with a shorter disease-free survival.

There are limited studies that describe the overexpression of Kpn $\beta$ 1 in cancer, although van der Watt et al. (2009) showed that Kpn $\beta$ 1 is expressed at elevated levels in cervical tumour tissue and cell lines compared to normal cervical epithelium (39), and Smith et al. (2010) found that Kpn $\beta$ 1 mRNA was elevated in ovarian cancer cell lines and transformed ovarian cells compared to normal primary ovarian epithelial cells (and to a certain extent in breast cancer cells compared to normal breast epithelial cells) (18). Furthermore, Kuusisto et al. (2011) described increased levels of Kpn $\beta$ 1 in several transformed cell lines, compared to their respective untransformed counterparts (32), suggesting that Kpn $\beta$ 1 upregulation is indeed associated with cellular transformation and cancer.

As the nuclear transport cycle employed by Karyopherin  $\beta$  proteins often requires energy provided by Ran, it follows that for increased Karyopherin  $\beta$  protein expression to translate into increased nuclear transport rates, Ran expression also needs to be elevated in cancer cells. This is in fact the case, as increased expression of Ran has been reported in various cancer types, with the cancer cell reported to be reliant on high levels of Ran expression for survival (53).

Based on these literature reports, we propose a model, shown in Figure 1, where increased expression of certain Karyopherin  $\beta$  proteins in cancer cells results in increased nuclear transport efficiency, thus facilitating increased oncogenic signalling and promoting the cancer phenotype.

### **Inhibition of Karyopherin $\beta$ proteins as a cancer therapy**

Since altered Karyopherin  $\beta$  protein expression and function has been shown to play a role in cancer development, the inhibition of Karyopherin  $\beta$  proteins could be an important anti-cancer strategy. However, drugs targeting these proteins are currently limited. Leptomycin B (LMB, also known as CI-940 or elactocin), a small molecule inhibitor of Crm1 (54), is an unsaturated branched-chain fatty acid that alkylates a single cysteine residue on the Crm1 protein (55), thereby preventing Crm1 from binding to and transporting its cargo proteins. It was shown to exhibit anti-cancer activity *in vitro* and *in vivo* (56) and was tested as a potential cancer therapeutic against a range of tumours in phase I clinical trials (57), although exhibited high toxicity when administered systemically, and so was discontinued

due to the associated side-effects (57). It is not clear whether these side-effects were due to off-target toxicity and many maintain that targeting Crm1 could still prove to be a novel and potent anti-cancer therapy, perhaps in lower concentrations or in combination with other therapies (58). Alternatively, a Crm1 inhibitor might be better tolerated if administered as a local rather than systemic therapy (59). Ultimately, LMB has established a model for inhibition of Crm1, and has also proved to be extremely useful for the analysis of nucleocytoplasmic transport of hundreds of endogenous, as well as viral proteins. LMB also acts as a reference point from which novel inhibitors can be designed. Mutka et al. (2009) has recently published a report on a series of new nuclear export inhibitors (NEIs) (60). These compounds are semisynthetic LMB derivatives that showed improved therapeutic potential compared to LMB. They maintain the high potency of LMB but are up to 16-fold better tolerated *in vivo* (60). Recent work has also reported the development of additional small molecules that target Crm1 and inhibit nuclear export, specifically N-azolyacrylates (61).

The identification of Crm1 as the protein target for LMB has spurred substantial interest in targeting the nuclear export and import machinery for drug discovery. Current work is underway in an attempt to “drug” other members of the Karyopherin  $\beta$  protein family, as well as to innovate creative ways of exploiting the nuclear transport process to improve the efficacy of current treatment options. For example, Aronov et al. (2004) describe the conjugation of anticancer agents, such as carboplatin analogues, to a poly(ethylene glycol) carrier and an NLS resulting in a rapid internalisation of the drug and an efficient accumulation in the nucleus (62). Although underused today, use of an NLS to direct drugs

to the nucleus may be a novel and efficient way to improve drug efficacy in the future (63). Another way in which the nuclear transport process could be exploited in order to improve the sensitivity of cancer cells to chemotherapeutic agents is by co-treating with a nuclear import inhibitor that can prevent the nuclear accumulation of proteins like NF $\kappa$ B and EGFR, whose respective nuclear accumulations are known to contribute to drug resistance following chemotherapy (64;65).

### **Kpn $\beta$ 1 as a target for cancer therapy?**

The increased expression and functional reliance on Kpn $\beta$ 1 in transformed and cancer cells suggests that targeting Kpn $\beta$ 1 itself may be an attractive approach to treating cancer. However, since both normal and cancer cells share the same nuclear transport machinery, there are concerns that its inhibition might induce side effects by inhibiting the proliferation of normal tissues. While this concern is valid, there is evidence that targeting proteins that normal cells require but on which cancer cells have increased dependence is a feasible cancer treatment option. An example is the targeting of Myc, a transcription factor required for the growth of somatic cells, yet to which cancer cells are “addicted” as it co-ordinates vital oncogenic signals (66). Soucek et al. (2008) showed that the inhibition of Myc *in vivo* triggered the rapid regression of lung tumours, and although it exerted effects on normal tissues, these effects were well tolerated and rapidly and completely reversible (67). Similar results have been obtained in other cancer types (66).

Following on from the success of targeting Myc in *in vivo* model systems, the targeting of Kpn $\beta$ 1 could have therapeutic benefits. This is evidenced in part by a study by van der Watt et al. (2009), which shows that the inhibition of Kpn $\beta$ 1 (using siRNA) in cancer and transformed cells leads to cell death via apoptosis, yet Kpn $\beta$ 1 inhibition in normal cells has only a minor effect on cell viability (39). Thus, the feasibility of targeting Kpn $\beta$ 1 as an anti-cancer therapeutic strategy needs to be evaluated. However, the limitation currently lies in the lack of availability of Kpn $\beta$ 1 inhibitors, or actually of nuclear import inhibitors in general. Up until a few years ago, the only available inhibitor of nuclear import was the lectin, wheat germ agglutinin (WGA) (68). WGA functions by blocking nuclear pore complexes, thus preventing nuclear import from occurring (69). Recently, however, a few nuclear import inhibitors with more specific targeting functions have been identified. These include two peptide inhibitors, bimax1 and bimax2, that were recently found to bind tightly to Karyopherin  $\alpha$  independently of Kpn $\beta$ 1, preventing the release of the cargo in the nucleus (70); and a peptide designated M9M, which was found to bind Karyopherin  $\beta$ 2 (transportin), inhibiting its nuclear import function (71). Small molecule peptidomimetic inhibitors of Karyopherin  $\alpha/\beta$  have also been described and used to study nuclear import *in vivo* (72), however, these inhibitors are not cell-permeable. Ivermectin is a broad-spectrum anti-parasitic recently found to inhibit Karyopherin  $\alpha/\beta$ , however, does not appear to block import mediated by Kpn $\beta$ 1 alone (73). Finally, Karyostatin 1A is the first novel small molecule inhibitor of Kpn $\beta$ 1 to be identified; however, its off-target effects have not yet been examined (74). In addition, it has not yet been determined whether Karyostatin 1A exhibits any anti-cancer effects. Overall, drugs targeting Kpn $\beta$ 1-mediated nuclear import are very limited, and much work remains to be done in order to determine the potential of Kpn $\beta$ 1, and its protein family in general, as a target for cancer therapy.

## **Conclusions and Perspectives**

The extensive roles played by Kpn $\beta$ 1 implicate it as a global regulator of the cell and essential for cell functioning. Yet, its deregulation in transformed and cancer cells and the associated increased reliance on its expression suggest that it has potential as a novel target for cancer therapy. Further research is required to fully elucidate the potential clinical usefulness of Kpn $\beta$ 1, and its other Karyopherin  $\beta$  family members, as anti-cancer therapeutic targets.

## **Acknowledgements**

This work is supported by funding from the Cancer Association of South Africa (CANSAs), the Medical Research Council (SA) and the University of Cape Town. Pauline van der Watt is supported by the Claude Leon Foundation Postdoctoral Fellowship Program.

## Figure legend

Figure 1. A diagrammatic representation of Karyopherin  $\beta$ -mediated nuclear transport in normal (a) and cancer (b) cells. a). Under normal conditions, nuclear import and export occurs predominantly via the Karyopherin  $\beta$  proteins, Kpn $\beta$ 1 and Crm1, respectively, where Kpn $\beta$ 1 can bind its cargoes directly, or in complex with its adaptor, Kpn $\alpha$ . b). In cancer cells, expression of certain Karyopherin  $\beta$  proteins (including Kpn $\beta$ 1 and Crm1) is

## References

- (1) Gorlich D, Kutay U. Transport between the cell nucleus and the cytoplasm. *Annu Rev Cell Dev Biol* 1999;15:607-60.
- (2) Harel A, Forbes DJ. Importin beta: conducting a much larger cellular symphony. *Mol Cell* 2004 Nov 5;16(3):319-30.
- (3) Fagotto F, Gluck U, Gumbiner BM. Nuclear localization signal-independent and importin/karyopherin-independent nuclear import of beta-catenin. *Curr Biol* 1998 Feb 12;8(4):181-90.
- (4) Moroianu J. Distinct nuclear import and export pathways mediated by members of the karyopherin beta family. *J Cell Biochem* 1998 Aug 1;70(2):231-9.
- (5) Kutay U, Izaurralde E, Bischoff FR, Mattaj JW, Gorlich D. Dominant-negative mutants of importin-beta block multiple pathways of import and export through the nuclear pore complex. *EMBO J* 1997 Mar 17;16(6):1153-63.
- (6) Friedrich B, Quensel C, Sommer T, Hartmann E, Kohler M. Nuclear localization signal and protein context both mediate importin alpha specificity of nuclear import substrates. *Mol Cell Biol* 2006 Dec;26(23):8697-709.
- (7) Kohler M, Speck C, Christiansen M, Bischoff FR, Prehn S, Haller H, et al. Evidence for distinct substrate specificities of importin alpha family members in nuclear protein import. *Mol Cell Biol* 1999 Nov;19(11):7782-91.
- (8) Lam MH, Briggs LJ, Hu W, Martin TJ, Gillespie MT, Jans DA. Importin beta recognizes parathyroid hormone-related protein with high affinity and mediates its nuclear import in the absence of importin alpha. *J Biol Chem* 1999 Mar 12;274(11):7391-8.
- (9) Lee SJ, Sekimoto T, Yamashita E, Nagoshi E, Nakagawa A, Imamoto N, et al. The structure of importin-beta bound to SREBP-2: nuclear import of a transcription factor. *Science* 2003 Nov 28;302(5650):1571-5.
- (10) Wychowski C, van der WS, Girard M. Nuclear localization of poliovirus capsid polypeptide VP1 expressed as a fusion protein with SV40-VP1. *Gene* 1985;37(1-3):63-71.
- (11) Dingwall C, Laskey RA. Nuclear targeting sequences--a consensus? *Trends Biochem Sci* 1991 Dec;16(12):478-81.
- (12) Chook YM, Blobel G. Karyopherins and nuclear import. *Curr Opin Struct Biol* 2001 Dec;11(6):703-15.
- (13) Ribbeck K, Lipowsky G, Kent HM, Stewart M, Gorlich D. NTF2 mediates nuclear import of Ran. *EMBO J* 1998 Nov 16;17(22):6587-98.
- (14) Strom AC, Weis K. Importin-beta-like nuclear transport receptors. *Genome Biol* 2001;2(6):REVIEWS3008.

- (15) Bednenko J, Cingolani G, Gerace L. Importin beta contains a COOH-terminal nucleoporin binding region important for nuclear transport. *J Cell Biol* 2003 Aug 4;162(3):391-401.
- (16) Beg AA, Ruben SM, Scheinman RI, Haskill S, Rosen CA, Baldwin AS, Jr. I kappa B interacts with the nuclear localization sequences of the subunits of NF-kappa B: a mechanism for cytoplasmic retention. *Genes Dev* 1992 Oct;6(10):1899-913.
- (17) Ushijima R, Sakaguchi N, Kano A, Maruyama A, Miyamoto Y, Sekimoto T, et al. Extracellular signal-dependent nuclear import of STAT3 is mediated by various importin alphas. *Biochem Biophys Res Commun* 2005 May 13;330(3):880-6.
- (18) Smith ER, Cai KQ, Smedberg JL, Ribeiro MM, Rula ME, Slater C, et al. Nuclear entry of activated MAPK is restricted in primary ovarian and mammary epithelial cells. *PLoS One* 2010;5(2):e9295.
- (19) Takizawa CG, Weis K, Morgan DO. Ran-independent nuclear import of cyclin B1-Cdc2 by importin beta. *Proc Natl Acad Sci U S A* 1999 Jul 6;96(14):7938-43.
- (20) Ciciarello M, Mangiacasale R, Thibier C, Guarguaglini G, Marchetti E, Di FB, et al. Importin beta is transported to spindle poles during mitosis and regulates Ran-dependent spindle assembly factors in mammalian cells. *J Cell Sci* 2004 Dec 15;117(Pt 26):6511-22.
- (21) Harel A, Chan RC, Lachish-Zalait A, Zimmerman E, Elbaum M, Forbes DJ. Importin beta negatively regulates nuclear membrane fusion and nuclear pore complex assembly. *Mol Biol Cell* 2003 Nov;14(11):4387-96.
- (22) Bauerle M, Doenecke D, Albig W. The requirement of H1 histones for a heterodimeric nuclear import receptor. *J Biol Chem* 2002 Sep 6;277(36):32480-9.
- (23) Pemberton LF, Paschal BM. Mechanisms of receptor-mediated nuclear import and nuclear export. *Traffic* 2005 Mar;6(3):187-98.
- (24) Kodiha M, Chu A, Matusiewicz N, Stochaj U. Multiple mechanisms promote the inhibition of classical nuclear import upon exposure to severe oxidative stress. *Cell Death Differ* 2004 Aug;11(8):862-74.
- (25) Spellman PT, Sherlock G, Zhang MQ, Iyer VR, Anders K, Eisen MB, et al. Comprehensive identification of cell cycle-regulated genes of the yeast *Saccharomyces cerevisiae* by microarray hybridization. *Mol Biol Cell* 1998 Dec;9(12):3273-97.
- (26) Quan Y, Ji ZL, Wang X, Tartakoff AM, Tao T. Evolutionary and transcriptional analysis of karyopherin beta superfamily proteins. *Mol Cell Proteomics* 2008 Jul;7(7):1254-69.
- (27) van der Watt PJ, Ngarande E, Leaner VD. Overexpression of Kpnbeta1 and Kpnalpha2 Importin Proteins in Cancer Derives from Deregulated E2F Activity. *PLoS One* 2011;6(11):e27723.
- (28) van der Watt PJ, Leaner VD. The nuclear exporter, Crm1, is regulated by NFY and Sp1 in cancer cells and repressed by p53 in response to DNA damage. *Biochim Biophys Acta* 2011 Jul;1809(7):316-26.
- (29) Yang W, Musser SM. Nuclear import time and transport efficiency depend on importin beta concentration. *J Cell Biol* 2006 Sep 25;174(7):951-61.

- (30) Feldherr C, Cole C, Lanford RE, Akin D. The effects of SV40 large T antigen and p53 on nuclear transport capacity in BALB/c 3T3 cells. *Exp Cell Res* 1994 Jul;213(1):164-71.
- (31) Feldherr CM, Akin D. Signal-mediated nuclear transport in proliferating and growth-arrested BALB/c 3T3 cells. *J Cell Biol* 1991 Nov;115(4):933-9.
- (32) Kuusisto HV, Wagstaff KM, Alvisi G, Roth DM, Jans DA. Global enhancement of nuclear localization-dependent nuclear transport in transformed cells. *FASEB J* 2011 Dec 12.
- (33) Behrens P, Brinkmann U, Fogt F, Wernert N, Wellmann A. Implication of the proliferation and apoptosis associated CSE1L/CAS gene for breast cancer development. *Anticancer Res* 2001 Jul;21(4A):2413-7.
- (34) Boni R, Wellmann A, Man YG, Hofbauer G, Brinkmann U. Expression of the proliferation and apoptosis-associated CAS protein in benign and malignant cutaneous melanocytic lesions. *Am J Dermatopathol* 1999 Apr;21(2):125-8.
- (35) Brinkmann U. CAS, the human homologue of the yeast chromosome-segregation gene CSE1, in proliferation, apoptosis, and cancer. *Am J Hum Genet* 1998 Mar;62(3):509-13.
- (36) Poon IK, Jans DA. Regulation of nuclear transport: central role in development and transformation? *Traffic* 2005 Mar;6(3):173-86.
- (37) Wellmann A, Krenacs L, Fest T, Scherf U, Pastan I, Raffeld M, et al. Localization of the cell proliferation and apoptosis-associated CAS protein in lymphoid neoplasms. *Am J Pathol* 1997 Jan;150(1):25-30.
- (38) Wellmann A, Flemming P, Behrens P, Wuppermann K, Lang H, Oldhafer K, et al. High expression of the proliferation and apoptosis associated CSE1L/CAS gene in hepatitis and liver neoplasms: correlation with tumor progression. *Int J Mol Med* 2001 May;7(5):489-94.
- (39) van der Watt PJ, Maske CP, Hendricks DT, Parker MI, Denny L, Govender D, et al. The Karyopherin proteins, Crm1 and Karyopherin beta1, are overexpressed in cervical cancer and are critical for cancer cell survival and proliferation. *Int J Cancer* 2009 Apr 15;124(8):1829-40.
- (40) Noske A, Weichert W, Niesporek S, Roske A, Buckendahl AC, Koch I, et al. Expression of the nuclear export protein chromosomal region maintenance/exportin 1/Xpo1 is a prognostic factor in human ovarian cancer. *Cancer* 2008 Apr 15;112(8):1733-43.
- (41) Yao Y, Dong Y, Lin F, Zhao H, Shen Z, Chen P, et al. The expression of CRM1 is associated with prognosis in human osteosarcoma. *Oncol Rep* 2009 Jan;21(1):229-35.
- (42) Shen A, Wang Y, Zhao Y, Zou L, Sun L, Cheng C. Expression of CRM1 in human gliomas and its significance in p27 expression and clinical prognosis. *Neurosurgery* 2009 Jul;65(1):153-9.
- (43) Huang WY, Yue L, Qiu WS, Wang LW, Zhou XH, Sun YJ. Prognostic value of CRM1 in pancreas cancer. *Clin Invest Med* 2009;32(6):E315.
- (44) Winnepenninckx V, Lazar V, Michiels S, Dessen P, Stas M, Alonso SR, et al. Gene expression profiling of primary cutaneous melanoma and clinical outcome. *J Natl Cancer Inst* 2006 Apr 5;98(7):472-82.

- (45) Dahl E, Kristiansen G, Gottlob K, Klaman I, Ebner E, Hinzmann B, et al. Molecular profiling of laser-microdissected matched tumor and normal breast tissue identifies karyopherin alpha2 as a potential novel prognostic marker in breast cancer. *Clin Cancer Res* 2006 Jul 1;12(13):3950-60.
- (46) Gluz O, Wild P, Meiler R, allo-Danebrock R, Ting E, Mohrmann S, et al. Nuclear karyopherin alpha2 expression predicts poor survival in patients with advanced breast cancer irrespective of treatment intensity. *Int J Cancer* 2008 Sep 15;123(6):1433-8.
- (47) Sakai M, Sohda M, Miyazaki T, Suzuki S, Sano A, Tanaka N, et al. Significance of karyopherin- $\alpha$  2 (KPNA2) expression in esophageal squamous cell carcinoma. *Anticancer Res* 2010 Mar;30(3):851-6.
- (48) Zheng M, Tang L, Huang L, Ding H, Liao WT, Zeng MS, et al. Overexpression of karyopherin-2 in epithelial ovarian cancer and correlation with poor prognosis. *Obstet Gynecol* 2010 Oct;116(4):884-91.
- (49) Wang CI, Wang CL, Wang CW, Chen CD, Wu CC, Liang Y, et al. Importin subunit alpha-2 is identified as a potential biomarker for non-small cell lung cancer by integration of the cancer cell secretome and tissue transcriptome. *Int J Cancer* 2011 May 15;128(10):2364-72.
- (50) Mortezaei A, Hermanns T, Seifert HH, Baumgartner MK, Provenzano M, Sulser T, et al. KPNA2 expression is an independent adverse predictor of biochemical recurrence after radical prostatectomy. *Clin Cancer Res* 2011 Mar 1;17(5):1111-21.
- (51) Jensen JB, Munksgaard PP, Sorensen CM, Frstrup N, Birkenkamp-Demtroder K, Ulhoi BP, et al. High expression of karyopherin-alpha2 defines poor prognosis in non-muscle-invasive bladder cancer and in patients with invasive bladder cancer undergoing radical cystectomy. *Eur Urol* 2011 May;59(5):841-8.
- (52) Yoshitake K, Tanaka S, Mogushi K, Aihara A, Murakata A, Matsumura S, et al. Importin-alpha1 as a novel prognostic target for hepatocellular carcinoma. *Ann Surg Oncol* 2011 Jul;18(7):2093-103.
- (53) Xia F, Lee CW, Altieri DC. Tumor cell dependence on Ran-GTP-directed mitosis. *Cancer Res* 2008 Mar 15;68(6):1826-33.
- (54) Nishi K, Yoshida M, Fujiwara D, Nishikawa M, Horinouchi S, Beppu T. Leptomycin B targets a regulatory cascade of crm1, a fission yeast nuclear protein, involved in control of higher order chromosome structure and gene expression. *J Biol Chem* 1994 Mar 4;269(9):6320-4.
- (55) Kudo N, Matsumori N, Taoka H, Fujiwara D, Schreiner EP, Wolff B, et al. Leptomycin B inactivates CRM1/exportin 1 by covalent modification at a cysteine residue in the central conserved region. *Proc Natl Acad Sci U S A* 1999 Aug 3;96(16):9112-7.
- (56) Roberts BJ, Hamelshle KL, Sebolt JS, Leopold WR. In vivo and in vitro anticancer activity of the structurally novel and highly potent antibiotic CI-940 and its hydroxy analog (PD 114,721). *Cancer Chemother Pharmacol* 1986;16(2):95-101.
- (57) Newlands ES, Rustin GJ, Brampton MH. Phase I trial of elactocin. *Br J Cancer* 1996 Aug;74(4):648-9.

- (58) Naniwa J, Kigawa J, Akeshima R, Kanamori Y, Itamochi H, Oishi T, et al. Leptomycin B enhances CDDP-sensitivity via nuclear accumulation of p53 protein in HPV-positive cells. *Cancer Sci* 2003 Dec;94(12):1099-103.
- (59) Gray LJ, Bjelogrić P, Appleyard VC, Thompson AM, Jolly CE, Lain S, et al. Selective induction of apoptosis by leptomycin B in keratinocytes expressing HPV oncogenes. *Int J Cancer* 2007 Jun 1;120(11):2317-24.
- (60) Mutka SC, Yang WQ, Dong SD, Ward SL, Craig DA, Timmermans PB, et al. Identification of nuclear export inhibitors with potent anticancer activity in vivo. *Cancer Res* 2009 Jan 15;69(2):510-7.
- (61) Van NT, Pannecouque C, Vanstreels E, Stevens M, Dehaen W, Daelemans D. Inhibition of the CRM1-mediated nucleocytoplasmic transport by N-azolyacrylates: structure-activity relationship and mechanism of action. *Bioorg Med Chem* 2008 Nov 1;16(21):9487-97.
- (62) Aronov O, Horowitz AT, Gabizon A, Fuertes MA, Perez JM, Gibson D. Nuclear localization signal-targeted poly(ethylene glycol) conjugates as potential carriers and nuclear localizing agents for carboplatin analogues. *Bioconjug Chem* 2004 Jul;15(4):814-23.
- (63) Chahine MN, Pierce GN. Therapeutic targeting of nuclear protein import in pathological cell conditions. *Pharmacol Rev* 2009 Sep;61(3):358-72.
- (64) Camp ER, Li J, Minnich DJ, Brank A, Moldawer LL, MacKay SL, et al. Inducible nuclear factor-kappaB activation contributes to chemotherapy resistance in gastric cancer. *J Am Coll Surg* 2004 Aug;199(2):249-58.
- (65) Huang WC, Chen YJ, Li LY, Wei YL, Hsu SC, Tsai SL, et al. Nuclear translocation of epidermal growth factor receptor by Akt-dependent phosphorylation enhances breast cancer-resistant protein expression in gefitinib-resistant cells. *J Biol Chem* 2011 Jun 10;286(23):20558-68.
- (66) Felsher DW. MYC Inactivation Elicits Oncogene Addiction through Both Tumor Cell-Intrinsic and Host-Dependent Mechanisms. *Genes Cancer* 2010 Jun;1(6):597-604.
- (67) Soucek L, Whitfield J, Martins CP, Finch AJ, Murphy DJ, Sodir NM, et al. Modelling Myc inhibition as a cancer therapy. *Nature* 2008 Oct 2;455(7213):679-83.
- (68) Yoneda Y, Imamoto-Sonobe N, Yamaizumi M, Uchida T. Reversible inhibition of protein import into the nucleus by wheat germ agglutinin injected into cultured cells. *Exp Cell Res* 1987 Dec;173(2):586-95.
- (69) Finlay DR, Newmeyer DD, Price TM, Forbes DJ. Inhibition of in vitro nuclear transport by a lectin that binds to nuclear pores. *J Cell Biol* 1987 Feb;104(2):189-200.
- (70) Kosugi S, Hasebe M, Entani T, Takayama S, Tomita M, Yanagawa H. Design of peptide inhibitors for the importin alpha/beta nuclear import pathway by activity-based profiling. *Chem Biol* 2008 Sep 22;15(9):940-9.
- (71) Cansizoglu AE, Lee BJ, Zhang ZC, Fontoura BM, Chook YM. Structure-based design of a pathway-specific nuclear import inhibitor. *Nat Struct Mol Biol* 2007 May;14(5):452-4.

- (72) Ambrus G, Whitby LR, Singer EL, Trott O, Choi E, Olson AJ, et al. Small molecule peptidomimetic inhibitors of importin alpha/beta mediated nuclear transport. *Bioorg Med Chem* 2010 Nov 1;18(21):7611-20.
- (73) Wagstaff KM, Rawlinson SM, Hearps AC, Jans DA. An AlphaScreen(R)-based assay for high-throughput screening for specific inhibitors of nuclear import. *J Biomol Screen* 2011 Feb;16(2):192-200.
- (74) Hintersteiner M, Ambrus G, Bednenko J, Schmied M, Knox AJ, Meisner NC, et al. Identification of a small molecule inhibitor of importin beta mediated nuclear import by confocal on-bead screening of tagged one-bead one-compound libraries. *ACS Chem Biol* 2010 Oct 15;5(10):967-79.

Figure 1

